§ 556.739 Trenbolone.

(a) *Acceptable daily intake (ADI).* The ADI for total residues of trenbolone is 0.4 microgram per kilogram of body weight per day.

(b) *Tolerances.* A tolerance for total trenbolone residues in uncooked edible tissues of cattle is not needed.

[64 FR 18574, Apr. 15, 1999]

§ 556.740 Tylosin.

Tolerances are established for residues of tylosin in edible products of animals as follows:

(a) In chickens and turkeys: 0.2 part per million (negligible residue) in uncooked fat, muscle, liver, and kidney.

(b) In cattle: 0.2 part per million (negligible residue) in uncooked fat, muscle, liver, and kidney.

(c) In swine: 0.2 part per million (negligible residue) in uncooked fat, muscle, liver, and kidney.

(d) In milk: 0.05 part per million (negligible residue).

(e) In eggs: 0.2 part per million (negligible residue).

§ 556.741 Tripelennamine.

A tolerance of 200 parts per billion (ppb) is established for residues of tripelennamine in uncooked edible tissues of cattle and 20 ppb in milk.

[62 FR 4164, Jan. 29, 1997]

§ 556.750 Virginiamycin.

(a) *Acceptable daily intake (ADI).* The ADI for total residues of virginiamycin is 250 micrograms per kilogram of body weight per day.

(b) *Tolerances*—(1) *Swine.* Tolerances are established for residues of virginiamycin in uncooked edible tissues of 0.4 part per million (ppm) in kidney, skin, and fat, 0.3 ppm in liver, and 0.1 ppm in muscle.

(2) *Broiler chickens and cattle.* A tolerance for residues of virginiamycin is not required.

[64 FR 48296, Sept. 3, 1999]

§ 556.760 Zeranol.

(a) *Cattle.* A tolerance for total zeranol residues in uncooked edible tissues of cattle is not needed. The safe concentration for total zeranol residues in uncooked edible tissues of cattle is 150 parts per billion (ppb) in muscle, 300 ppb in liver, 450 ppb in kidney, and 600 ppb in fat. A tolerance refers to the concentration of marker residues in the target tissue used to monitor for total drug residues in the target animal. A safe concentration refers to the total residue concentration considered safe in edible tissues.

(b) *Sheep.* No residues of zeranol may be found in the uncooked edible tissues of sheep as determined by the following method of analysis:

I. METHOD OF ANALYSIS—ZERANOL

A gas chromatographic method for the determination of the drug in frozen beef tissues is described. Tissue is frozen and stored in a deep freezer until ready for examination. A weighed portion of wet tissue (with exception of fat) is homogenized and lyophilized to dry solid. The drug is recovered from dry tissue by an extraction with methanol in a Soxhlet extractor. The methanol extract is digested in the presence of hydrochloric acid to hydrolyze conjugates should any be present. Elimination of impurities is brought about by liquid partition transfer successively to chloroform to 1*N* sodium hydroxide, to carbon tetrachloride, to 1*N* sodium hydroxide, to ethyl ether, and, finally, to a dry residue. The residue is reacted with a silane mixture to create a volatile derivative which is quantitated by peak area measurements from a flame ionization detector. The drug can be detected at a level of 20 parts per billion with negligible interference from tissues or reagents.

II. REAGENTS

A. Carbon tetrachloride, N.F., Fisher Scientific C–186, or equivalent.

B. Chloroform, N.F., Fisher Scientific C–296, or equivalent.

C. Chromatograph gases, flow rates adjusted to maximize sensitivity for specific chromatograph.

1. Carrier gas, conventional tank helium.

2. Flame makeup gas.

a. Oxygen, conventional tank oxygen.

b. Hydrogen, Linde high purity, or equivalent.

D. Column packing, 3 percent GE SE–52 (Applied Science Laboratories) on P.E. Celite 60–80 mesh (Johns Manville Product No. 154–0048), or equivalent.

E. Ether, anhydrous, Fisher Scientific E–138, or equivalent.

F. Hexamethyldisilazane, Dow-Corning, Peninsular, or equivalent.

G. Hydrochloric acid, analytical reagent grade.

H. Methanol, certified A.C.S., spectranalyzed, Fisher Scientific A–408, or equivalent.

I. Phosphoric acid, analytical reagent grade.

J. Pyridine, anhydrous, A.C.S. reagent grade.

K. Silating reagent mixture: Pipet 8 milliliters each of pyridine and hexamethyldisilazane and 4 milliliters of trimethylchlorosilane into a clean glass vial with a polyethylene cap and mix thoroughly. Let stand overnight and decant supernatant liquid into a vial. Cap and store at room temperature for daily use. If kept dry, the reagent is stable for more than a month. Blanks are scanned by gas chromatography on each new bottle of J, F, and N material used in the silating reagent mixture for possible peak interference in the region of zeranol derivative.

L. Sodium chloride, analytical reagent grade.

M. Sodium hydroxide, analytical reagent grade.

N. Trimethylchlorosilane, Dow-Corning, Peninsular, or equivalent.

O. Water, distilled in glass.

P. Zeranol, primary standard.

Q. Solutions.

1. 2*N* Hydrochloric acid in water.

2. 3*N* Phosphoric acid in water.

3. 2 percent w/v solium chloride in water.

4. 1*N* Sodium hydroxide in water.

III. Apparatus

A. Extraction assemblies, Soxhlet, improved, standard taper grindings, Pyrex brand glass, 1,000 milliliters capacity, Sargent Catalog S–31265D, or equivalent.

B. Flasks, freeze drying, widemouth, 1,000 milliliters capacity, 24/40 standard taper grindings, Pyrex brand glass, Sargent Catalog S–28875–20–F, or equivalent.

C. Flasks, homogenizing, 250 milliliters, Sargent Catalog S–61716, or equivalent.

D. Funnels, separatory, Squibb stopper, with Teflon stopcock plug, Pyrex brand glass, 250- and 500-milliliter capacities, Sargent Catalog S–35815–20–F or G, or equivalent.

E. Gas chromatograph, F and M Model 5750 with flame ionization detector, or equivalent.

F. Gas chromatography column: Stainless steel tubing, 6 feet by 3⁄16 inch packed with 3 percent by weight GE SE–52 (Applied Science Laboratories) deposited on P.E. Celite 60–80 mesh (product No. 154–0048), or equivalent. Condition the column by baking for 40–80 hours at 325 °C. with a helium flow, but detached from the detector input. Injections of 1–2 microliters of a 50/50 mixture of hexamethyldisilazane and trimethylchlorosilane will help remove active sites in the column.

1. Prepare a TMS derivative of a 1,000-microgram zeranol standard as described in the procedure section. Inject 1-microliter quantities to determine whether the column is responding to the conditioning. After the column shows a response at the 1,000-microgram level, proceed to smaller quantities to optimize conditions.

2. The column and chromatograph must be conditioned to achieve a minimum sensitivity response so that a peak 5 millimeters in height results from an injection of 5 microliter of standard preparation containing 1 microgram of zeranol in the derivative preparation. This criterion must be met before tissue assay is attempted.

3. The column is brought to 250 °C. after conditioning and held at that temperature for at least 12 hours before making a run.

G. Heating mantle, electric, Glas-Col. Sargent Catalog S–40866H, or equivalent.

H. Hot plate, with gradient rheostat heat control.

I. Meat grinder, manually operated or equivalent.

J. Steam bath.

K. Syringe, Hamilton Micro Syringe Model 701, 10-microliter capacity, or equivalent.

L. Torsion balance, 0.1 gram sensitivity, 500 grams capacity.

M. Vials, 1-dram glass with plastic tops, Owens-Illinois, Opticlear, or equivalent.

N. Virtis freeze drier, Sargent Catalog S–28881–80, or equivalent.

O. Virtis homogenizing mill, macro, Virtis No. 45, Sargent Catalog S–61700, or equivalent.

IV. Standard Solutions

A. Stock solution A: Accurately weigh 0.1000 gram of zeranol, primary standard, into a 250-milliliter beaker. Dissolve the standard in 80 milliliters of methanol and accurately dilute to 100 milliliters in a volumetric flask with methanol. By preparation, the solution contains 1,000 micrograms per milliliter.

B. Stock solution B: Dilute 10.0 milliliters of stock solution A to 100 milliliters with methanol to provide a standard containing 100 micrograms of the drug per milliliter.

C. Stock solution C: Dilute 5.0 milliliters of stock solution B to 100 milliliters with methanol to provide a standard of 5 micrograms per milliliter.

D. Stock solution D: Dilute 2.0 milliliters of stock solution B to 100 milliliters with methanol to provide a standard of 2 micrograms per milliliter. Transfer 1.0 milliliter of stock solution D to a 1-dram glass vial, evaporate to a dry residue in a vacuum desiccator at reduced pressure. The residue contains 2 micrograms of zeranol to be used as a calibration standard in operation of the gas chromatograph.

V. PROCEDURE

A. Preparation of glassware: Glassware should be washed in detergent or chromic acid solution to remove contaminants and rinsed in water to remove traces of cleaning agent. Rinse with methanol before using.

B. Preparation of sample.

1. Collect muscle, liver, kidney, and tripe from a freshly sacrificed animal under the cleanest conditions possible.

2. Grind the fresh tissue in a meat grinder, divide into 100-gram portions, and wrap in aluminum foil. Store wrapped tissue in a deep freeze. Fat should be wrapped in foil and stored in deep freeze.

C. Extraction procedure for muscle, liver, kidney, and tripe.

1. Weight 100 grams of partially thawed tissue into a 250-milliliter homogenizing flask, add 60 milliliters of water, and attach to a Virtis ''45'' Tissue Mill, or equivalent.

2. Mix the materials at 45,000 r.p.m. for 5 minutes to obtain a thin homogenate.

3. Transfer the homogenate to a 1-liter, widemouth, freeze drying flask using 10–20 milliliters of water for a rinse.

4. Place the flask on its side in a nearly horizontal position in a slurry of dry ice and acetone. Rotate the flask on its side as the homogenate cools to set down a uniform frozen solid layer on the wall of the flask.

5. Mount the flask on a Virtis freeze drier, or equivalent, and lyophilize to dry solids. This operation usually requires 20–24 hours. *Stopping place.*

6. Transfer the solid cake to a clean sheet of paper and crumble by hand to a size convenient for transfer to an extraction thimble.

7. Transfer the solids to a single thickness 60 x 180 milliliter Soxhlet extraction thimble and compact the solids sufficiently to guarantee complete immersion during solid extraction.

8. Transfer 600 milliliters of methanol to a 1-liter pot of a Soxhlet extraction assembly and place the thimble in the extractor. Mount a large glass funnel in the neck of the extractor with the stem extending into the thimble. Rinse the 1-liter freeze drying flask with three 50-milliliter portions of fresh methanol and transfer the rinses through the funnel into the thimble. Mount the condenser in the extractor and extract the solids for 15 hours. The extractor should be heated with the electric heating mantle so that a fill-empty cycle requires 18–24 minutes.

9. Drain the methanol from the thimble. Composite the methanol from the extractor and pot in an 800-milliliter beaker.

10. Rinse the pot with 10 milliliters of methanol and add to the methanol composite. Transfer 50 milliliters of 2*N* HCl down the pot side wall, and add to methanol composite. Concentrate to 125 milliliters by boiling on a hot plate.

D. Extraction procedure for fat.

1. Cut fat into ¼-inch cubes. The lyophilization of fat is unnecessary since it is essentially water free.

2. Transfer 100 grams of the prepared fat to a 60- x 180-millimeter extraction thimble and extract with 750 milliliters of methanol for 15 hours in the Soxhlet extractor. The extractor should be heated with the electric heating mantle so that a fill-empty cycle requires 18–24 minutes.

3. Drain the methanol from the thimble. Composite the methanol from the extractor and pot into an 800-milliliter beaker.

4. Rinse the pot with 10 milliliters of methanol and add to the methanol composite. Transfer 50 milliliters of 2*N* HCl down the pot side wall, and add to methanol composite. Concentrate to 125 milliliters by boiling on a hot plate.

E. Solvent partition.

1. Transfer the methanol concentrate to a 500-milliliter separatory funnel, identified by number as 1, with 70 milliliters of chloroform rinse and mix.

2. Add 300 milliliters of water and without shaking allow liquid phases to separate.

3. Withdraw the chloroform layer into a separatory funnel, identified by number as 2, containing 100 milliliters of 2 percent aqueous sodium chloride.

4. Gently mix the contents of funnel 2 horizontally end to end 30 times and allow phases to separate. Usually about 20 minutes are required to obtain maximum chloroform separation.

5. Withdraw the chloroform layer into a beaker.

6. Extract with shaking the contents of funnels 1 and 2 successively with three more 50-milliliter portions of chloroform.

7. Composite the chloroform extracts and concentrate to 125 milliliters by evaporation on a steam bath and cool to room temperature.

8. Transfer the chloroform composite to a 250-milliliter separatory funnel, fitted with a Teflon stopcock, using 10 milliliters of chloroform as a rinse.

9. Extract the chloroform with three separate 20-milliliter portions of 1*N* sodium hydroxide solution retaining the emulsion in the sodium hydroxide phase. Agitation of sodium hydroxide with the chloroform extract for the first time is accompanied by the appearance of emulsion.

10. Perform an extraction by gently inverting the closed funnel and returning the funnel to an upright position.

11. Repeat phase mixing 30 times per extraction.

12. Allow phases to separate for 10 minutes. The time delay allows for gradual dissipation of the emulsion to improve phase separation. The zeranol transfers from the chloroform to the upper sodium hydroxide phase in this operation.

13. Composite the sodium hydroxide extracts.

14. Wash the sodium hydroxide extract with three 50-milliliter portions of chloroform using the technique as in step 9 and the same 10-minute interval for phase separation. Washing the chloroform removes the emulsion and unwanted impurities from the sodium hydroxide phase.

15. Discard the chloroform washes. Transfer the sodium hydroxide extracts to a 250-milliliter beaker. Rinse each separatory funnel with two 5-milliliter portions of water and add to the sodium hydroxide extract. Wash each funnel twice with tap water and twice with distilled water before next use.

16. Neutralize the washed sodium hydroxide extract to pH 8.0 by dropwise addition of 3*N* phosphoric acid using a pH meter for pH detection.

17. Transfer the pH 8.0 water extract to a 250-milliliter separatory funnel using 10 to 20 milliliters of water for a rinse.

18. Extract the solution with three separate 50-milliliter portions of carbon tetrachloride. The zeranol transfers to the lower carbon tetrachloride phase. Use the same 30-count phase-mixing technique as in step 9 and allow the mixture to stand 5 minutes for phase separation.

19. Composite the carbon tetrachloride extracts.

20. Extract the carbon tetrachloride composite with two 20-milliliter portions of 1*N* sodium hydroxide. Zeranol transfers from carbon tetrachloride to the upper sodium hydroxide phase. After phase mixing, allow the mixture to stand 5 minutes for phase separation.

21. Composite the sodium hydroxide extracts.

22. Wash the extract with two 50-milliliter portions of carbon tetrachloride. Allow the mixture to stand 5 minutes for phase separation. Discard the carbon tetrachloride washes.

23. Transfer the sodium hydroxide extract into a 250-milliliter beaker. Rinse the separatory funnel with two 5-milliliter portions of water and add to the sodium hydroxide extract. Wash each funnel twice with tap water and twice with distilled water before next use. Adjust the sodium hydroxide extract to a pH of 9.5 by dropwise addition of 3*N* phosphoric acid and transfer to a 250-milliliter separatory funnel using 10–20 milliliters of water for a rinse.

24. Extract the pH 9.5 water solution with three separate 30-milliliter portions of anhydrous ethyl ether. Allow the mixture to stand 5 minutes for phase separation. The zeranol transfers to the upper ether phase.

25. Composite the ether extracts in a 125-milliliter Erlenmeyer flask.

26. Reduce the volume of ether to about 1–2 milliliters by evaporation on a hot plate with low heat while removing vapor from top of flask by vacuum aspiration.

27. Transfer ether residue to a 1-dram glass vial. Rinse down flask side wall with 1–2 milliliters of fresh ether and transfer to the glass vial.

28. Continue evaporation of ether to 0.1 milliliter.

29. Place vial in a vacuum desiccator and evaporate residue at line vacuum and room temperature overnight to dryness.

30. Close vial with a plastic cap and submit ether residue for preparation of TMS derivative and gas chromatographic assay. *Stopping place.*

F. Gas liquid chromatography.

1. Start the gas chromatography and maintain the following operational conditions:

Carrier gas pressure: 50 p.s.i. at tank.

Carrier gas flow rate: Sufficient to give zeranol derivative peak a retention time of 4–8 minutes.

Electrometer range: 10^2 or 10^1.

Detector temperature: 325 °C.

Injection port temperature: 325 °C.

Column temperature: 250°–280 °C., operate isothermally.

Recorder sensitivity: 1 millivolt.

Recorder chart speed: 1 inch per minute.

Sample size: 1 microliter to 5 microliters as necessary to give desired peak area for quantitative measurement.

Septums: Replace each evening and allow to condition overnight at operational temperature.

Flame assembly: Remove silica ash from the flame assembly each week. The flame assembly is removed; the anode, flame jet, and chimney are cleaned with a nylon bristle brush. Water and acetone are drawn through the jet capillary to remove any foreign material.

2. Add 0.2 milliliter of silating reagent to the sample or to the zeranol standard.

3. Stopper the vial and shake vigorously.

4. Warm the vial at 40°–50° C. for a few minutes, then roll the vial on a horizontal plane to insure that all of the interior surfaces of the vial have been in contact with the reagent.

5. Let vial stand for 4 hours or overnight in a warm area (40 °C.) to allow reaction to reach completion.

6. Place vial in a small padded centrifuge tube and centrifuge to settle the precipitate and insure that all the liquid is at the bottom of the vial.

7. Inject 1.0–5.0 microliters of clear solution into the chromatograph. At the beginning of the day's run, make 3–5 injections of a standard to condition the column for that day before taking quantitative data.

8. Run known mixtures at the beginning, middle, and end of the day's run over the concentration range of samples to be analyzed to compensate for day-to-day sensitivity fluctuations and drift. If four or less

samples are to be run, calibrating at the beginning and end of the run is sufficient.

VI. CALCULATIONS

Area values are obtained on known mixtures and samples by multiplying the net peak height by the peak width at half height or by counting squares. Area values obtained on knowns are plotted versus zeranol concentration. Calibration plots indicate a near linear function in the 0–10 microgram range. Area values obtained on samples are converted directly to microgram quantities using the curve. Control tests demonstrated a 70 percent recovery of zeranol from spiked wet beef liver and muscle necessitating a correction factor.

$$\text{Zeranol, parts per billion} = \frac{\text{Micrograms of zeranol found A1,000}}{\text{W A0.7}}$$

Where:

0.7=Correction factor for 70 percent recovery.

W=Grams of tissue examined.

VII. RECOVERY STUDY

A. Fortification of reagent blank.

1. For those using this method for the first time either for recovery study or tissue assay, a solvent blank and solvent fortified with zeranol should be processed through the entire procedure. This preliminary operation will establish whether or not the procedure is free from contamination arising from solvents and glassware and demonstrate the level of recovery of the standard zeranol. Level of recovery should be in the same range as the samples.

2. Transfer 600 milliliters of methanol to a 1-liter beaker. Add 50 milliliters of 2*N* HCl to the methanol and concentrate to 125 milliliters by boiling on a hot plate.

3. Transfer 600 milliliters of methanol to a 1-liter beaker. Add 50 milliliters of 2*N* HCl to the methanol and concentrate to 125 milliliters by boiling on a hot plate. Spike the concentrate with 1.0 milliliter of stock solution D.

4. Assay both samples as described in the procedure beginning extraction step V–E1.

B. Fortification of samples.

1. Transfer 100-gram portions of partially thawed tissues into 250-milliliter homogenizing flasks and set half of them aside to serve as tissue blanks.

2. Add to the remaining samples 1 milliliter of stock solution D to serve as fortified samples to which 20 parts per billion zearalanol have been added.

3. Assay both fortified and unfortified tissue as described in the procedure section beginning with V–C1.

[40 FR 13942, Mar. 27, 1975, as amended at 54 FR 31950, Aug. 3, 1989]

§ 556.770 Zoalene.

Tolerances are established for residues of zoalene (3,5-dinitro-*o*-toluamide) and its metabolite 3-amino-5-nitro-*o*-toluamide in food as follows:

(a) In edible tissues of chickens:

(1) 6 parts per million in uncooked liver and kidney.

(2) 3 parts per million in uncooked muscle tissue.

(3) 2 parts per million in uncooked fat.

(b) In edible tissues of turkeys: 3 parts per million in uncooked muscle tissue and liver.

PART 558—NEW ANIMAL DRUGS FOR USE IN ANIMAL FEEDS

Subpart A—General Provisions

Sec.

Subpart B—Specific New Animal Drugs For Use in Animal Feeds