

are not known to be infected with or exposed to tuberculosis may not be moved interstate only if they are accompanied by VS Form 1-27 and are moved interstate in an officially sealed means of conveyance directly to slaughter at an approved slaughtering establishment.

§ 77.32 General restrictions.

(a) Except for movement from accredited-free States and zones in accordance with § 77.23, movement from accredited herds in accordance with § 77.35, and movement to slaughter in accordance with §§ 77.25(a), 77.27(a), 77.29(a), and 77.31(d), no captive cervid may be moved interstate unless it has been tested using an official tuberculosis test, and it is moved in compliance with this part.

(b) No captive cervid with a response to any official tuberculosis test is eligible for interstate movement unless the captive cervid subsequently tests negative to a supplemental official tuberculosis test or is moved interstate directly to slaughter or necropsy in accordance with § 7.40.

(c) Except for captive cervids moving interstate under permit directly to slaughter or necropsy under § 77.40, each captive cervid or shipment of captive cervids to be moved interstate must be accompanied by a certificate issued within 30 days of the movement by a State or Federal animal health official or an accredited veterinarian.

(d) Captive cervids in zoological parks that have been accredited by the American Zoo and Aquarium Association (AZA) are exempt from the regulations in this part when the captive cervids are moved directly interstate between AZA member facilities. Any captive cervids moved interstate that are not moved directly from an AZA member facility to another AZA member facility must be moved in accordance with the regulations in this subpart.

§ 77.33 Testing procedures for tuberculosis in captive cervids.

(a) *Approved testers.* Except as explained in paragraphs (a)(1) and (a)(2) of this section, official tuberculosis tests may only be given by a veterinarian employed by the State in which

the test is administered or by a veterinarian employed by USDA.

(1) A designated accredited veterinarian may conduct the SCT test, except as provided in § 77.34(a)(2) and § 77.39(e) and (f).

(2) Any accredited veterinarian may conduct the BTB test.

(b) *Approved diagnostic laboratories.* (1) With one exception, histopathology and culture results for all tuberculosis diagnoses will be accepted only from the National Veterinary Services Laboratories (NVSL) in Ames, IA. The exception is that results will be accepted from a laboratory of the Food Safety and Inspection Service, USDA, for tissue examination of regular-kill slaughter animals in those cases where no submission is made to NVSL.

(2) The following laboratory is approved to perform the BTB test: Texas Veterinary Medical Center laboratory at Texas A&M University in College Station, TX.

(c) *Identification.* Any captive cervid tested with an official tuberculosis test must bear official identification in the form of an official eartag, or another identification device or method approved by the Administrator as unique and traceable, at the time of the official tuberculosis test. Use of any identification device or method other than an official eartag must first be approved by the Administrator as unique and traceable. Written requests for approval must be sent to National Animal Health Programs, VS, APHIS, 4700 River Road Unit 43, Riverdale, MD 20737-1231.

(d) *Reporting of tests*—(1) *SCT and CCT tests.* For the SCT and CCT tests, the testing veterinarian must submit a report to cooperating State and Federal animal health officials of the State in which the captive cervid is tested. The report must include the following information for all SCT and CCT tests administered: The number of the individual eartag or other identification approved by the Administrator; the age, sex, and breed of each captive cervid tested; a record of all responses; the size of each response for the CCT test; and the test interpretation.

(2) *BTB test.* Copies of the BTB test results must be submitted by the testing laboratory to the person, firm, or

corporation responsible for the management of the herd, cooperating State and Federal animal health officials of the State in which the captive cervid is tested, and the testing veterinarian. The report must include the following information for all BTB tests administered: The number of the individual eartag or other identification approved by the Administrator; the age, sex, and breed of each captive cervid tested; the test interpretation, and a summary of supporting data. Full supporting data must be submitted by the testing laboratory on a case-by-case basis at the request of cooperating State and Federal animal health officials.

(e) *Test interpretation.*(1) Interpretation of an SCT test will be based upon the judgment of the testing veterinarian after observation and palpation of the injection site, in accordance with the classification requirements described in § 77.34(a).

(2) Interpretation of a CCT test will be in accordance with the classification requirements described in § 77.34(b).

(3) Interpretation of a BTB test will be in accordance with the patented standards for the BTB test² and the classification requirements described in § 77.34(c).

(f) *Captive cervids eligible for testing.* Except as provided in § 77.35(a)(1) and § 77.36(a)(1), testing of herds for individual herd classification must include all captive cervids 1 year of age or over and any captive cervids other than natural additions (captive cervids born into the herd) under 1 year of age.

§ 77.34 Official tuberculosis tests.

(a) *Single cervical tuberculin (SCT) test.*

(1) The SCT test is the primary test to be used in individual captive cervids and in herds of unknown tuberculous status. Each captive cervid that responds to the SCT test must be classified as a suspect until it is retested with either the CCT test or the BTB

test and is either found negative for tuberculosis or is classified as a reactor, unless, with the exception of a designated accredited veterinarian, the testing veterinarian determines that the captive cervid should be classified as a reactor based on its response to the SCT test. A designated accredited veterinarian must classify a responding captive cervid as a suspect, unless the DTE determines, based on epidemiological evidence, that the captive cervid should be classified as a reactor.

(2) The SCT test is the primary test to be used in affected herds and in herds that have received captive cervids from an affected herd. When used with affected herds or in herds that have received captive cervids from an affected herd, the SCT test may only be administered by a veterinarian employed by the State in which the test is administered or employed by USDA. In affected herds or herds that have received captive cervids from an affected herd, each captive cervid that responds to the SCT test must be classified as a reactor, unless the DTE determines that the captive cervid should be classified as a suspect because of possible exposure to a tuberculous animal.

(b) *Comparative cervical tuberculin (CCT) test.* (1) The CCT test is a supplemental test that may only be used for retesting captive cervids classified as suspects. The CCT test may be used in affected herds only after the herd has tested negative to at least two whole herd SCT tests and only with the prior written consent of the DTE. The CCT test may not be used as a primary test for herds of unknown tuberculous status.

(2) A captive cervid tested with the CCT test must be classified as negative if it has a response to the bovine PPD tuberculin that is less than 1 mm.

(3) Unless the testing veterinarian determines that the captive cervid should be classified as a reactor because of possible exposure to a tuberculous animal, a captive cervid tested with the CCT test must be classified as a suspect if:

(i) It has a response to the bovine PPD tuberculin that is greater than 2 mm and that is equal to the response to the avian PPD tuberculin; or

²The patented standards for the BTB test may be obtained from the Texas Veterinary Medical Center, College of Veterinary Medicine, Texas A&M University, College Station, TX, or from the Deer Research Laboratory, Department of Microbiology, University of Otago, P.O. Box 56, Dunedin, New Zealand.